



The Beneficial Effects of Photobiomodulation to Reduce Intraocular Pressure in Primary Open-Angle Glaucoma

A. Chaoui Boudghene^{a,b*}, T. I. Braik Ahmed^a, K. Belbachir^a
and H. Benkazdali Ghalia^a

^a SARL Laboratory Neurogenesis Research and Development, Mostaganem, Algeria.

^b EURL Clinique Dar El Bassar, Mostaganem, Algeria.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/OR/2023/v18i3387

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/98246>

Original Research Article

Received: 17/02/2023

Accepted: 19/04/2023

Published: 17/05/2023

ABSTRACT

Purpose: The aim of this trail was to study the efficacy of photobiomodulation (PBM) treatment to reduce the intraocular pressure in subjects with primary open angle glaucoma disease.

Methods: Twenty eyes suffering from open angle glaucoma with high IOP level were selected, examined and treated with Thera-RED light diode system. The subjects were divided into two groups (n=10); treated and placebo group respectively; this system provide two lights (red at 660nm continue and Near IR light micro-pulsed at 850 nm), the patients received two series of treatment (ten per month within three months between every series of sessions) over five months and follow up taking ocular pressure measurements after every diode delivery session.

Results: A significant decrease in IOP has been observed from the first month of the treatment compared to the placebo group (from 22.6mmHg before treatment to 15 mmHg after the tenth

*Corresponding author: E-mail: amichab@yahoo.fr, amine.chaoui.boudghene@laboratoryneurogenesis.com;

session) this amelioration has been also remarked during the second session of treatment (fifth month) to attempt 14.2 mmHg.

Conclusion: Thera-RED light diode system treatment shows a significant decrease of IOP and remained to be stable under 14.2 mmHg in all the follow up, which confirm that photobiomodulation help in reducing the intraocular pressure in glaucomatous patients .

Keywords: Photobiomodulation; intra ocular pressure; trabecular meshwork; diode light; primary open angle glaucoma; aqueous humor.

1. INTRODUCTION

“Primary Open Angle Glaucoma (POAG) is a degenerative and chronic optic neuropathy” [1] “it is a complex, multifactorial neurodegenerative disease process that leads to progressive damage to the optic nerve [2], and the leading cause of irreversible blindness in the world” [1].

Up to date, glaucoma presents a number of cases about 76 million in 2020 and could reach 111.8 million in 2040 [3,4].

“The most important risk factor for glaucoma is the increase of intraocular pressure (IOP) [5], the IOP is generated by a damage of trabecular meshwork” [6].

“The use of photobiomodulation therapy has recently been considered in many diseases by stimulating cell migration and proliferation towards the damaged tissue, and controlling inflammation, which will eventually cure the disease” [7].

Our study is for the purpose of stabilized the intraocular pressure with ameliorating the aqueous humor outflow.

2. MATERIALS AND METHODS

20 eyes with primary open angle glaucoma (POAG) were selected and divided into 2 groups (n=10). The first group was treated with PBM; the second group was considered as placebo group. The subjects were selected after ophthalmological exams (VA, SL, gonioscopy) and no glaucoma treatment has been stopped during the study. This study was conducted in partnership between the laboratory neurogenesis research and development and ophthalmological single center.

2.1 Study Design

The data were collected during 20 visits over the course of 5 months' study (Fig. 1) no exclusion

factors were present; a written informed consent was taking from the patients. And the good practice of Helsinki guide lines was applied.

The clinical studies were carried out within an ophthalmological clinic located in Algeria.

The device has been patented and certified by the Algerian national institute of industrial property under the reference (N°230300/26/03/2023).

2.2 Exclusion Factors

Patients with Close angle glaucoma; glaucoma with normal intraocular pressure and Mono ophthalmic patients were excluded from this study.

Subjects underwent two rounds of treatment during the study which consisted of PBM or placebo treatments with the assurance of their anonymity, 10 sessions for 31 days (1 month) with an interval of three months maximum between the series of sessions initiated at the start and repeated in the 5th month.

The procedures involving the treatment of our patients were carried out under good hygienic and regulatory conditions, and good practice of Helsinki.

Intra ocular pressure (IOP) was taken before and after each session with aplanation tonometry and a diagram of the evolution of the ocular pressure was provided in order to establish the IOP evolution.

2.3 Evaluated Parameters

Tonometry by aplanation tests as well as a visual field with 24/2 program (Zeiss) were taken in order to establish a statistical study highlighting the results of our study.

2.4 Photobiomodulation Treatment

The subjects were treated with PBM “Thera-Red” which delivers two distinct wavelengths, a

red range (660 n.m) and a near IR range (850 n.m), with emitted energy of 300mW/cm² pulsed for the near IR, and an emitted energy of

250mW/cm² for red light. The treatment consists of projecting red and Near IR lights in 4 distinct phases under a well-established protocol (Table 1).

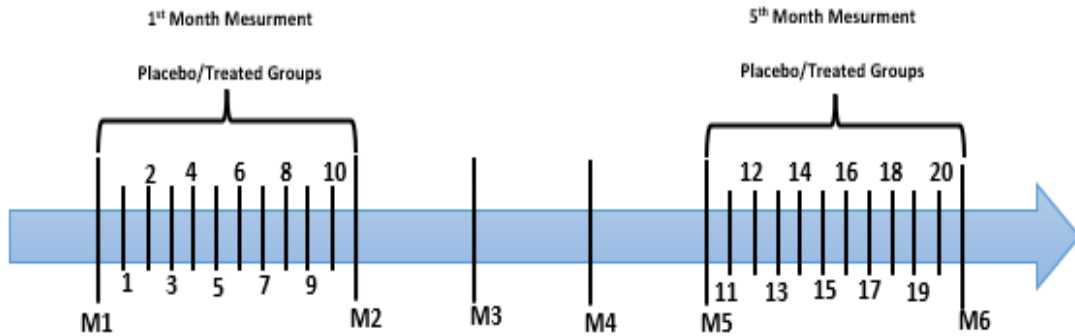


Fig. 1. Diagram illustrating the PBM clinical study design

Table 1. ‘Thera- Red’ system specifications

Parameter	Specifications
Light source	Diode light emission.
Light emission	660 n.m output 300mW/cm ² 850 n.m output 250mW/cm ²
Treatment exposure time	Total of 280 seconds (4 minutes 40 seconds). Dispatched into 4 phases: <ol style="list-style-type: none"> 1. 100 seconds with closed eye continuous RED wavelength. 2. 40 seconds with open eye pulsed Near IR wavelength. 3. 100 seconds with closed eye continuous RED wavelength. 4. 40 seconds with open eye pulsed Near IR wavelength.



Fig. 2. Thera-Red

The first phase includes the projection of continuous red diode light for 100 seconds (eye closed), followed by a second phase in which a pulsed near IR light is projected for 40 seconds (eye open). A third phase follows comprising continuous red light for 100 seconds (eye closed), and the fourth and final phase consists of a second projection of near IR light for 40 seconds (eye open).

2.5 Statistical Analyses

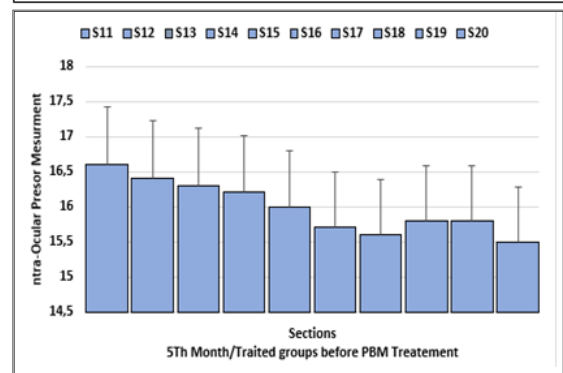
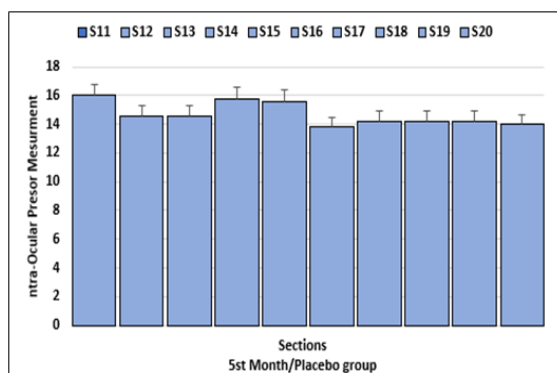
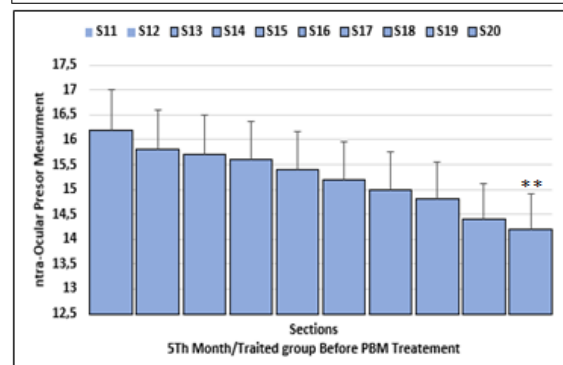
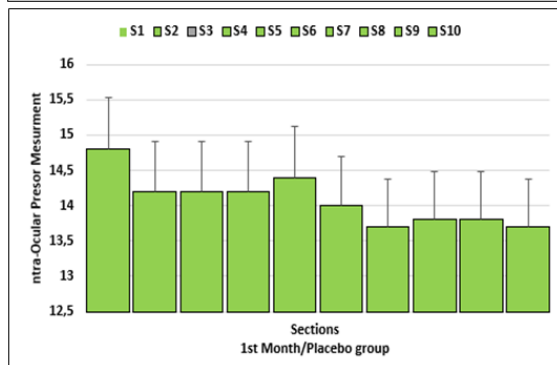
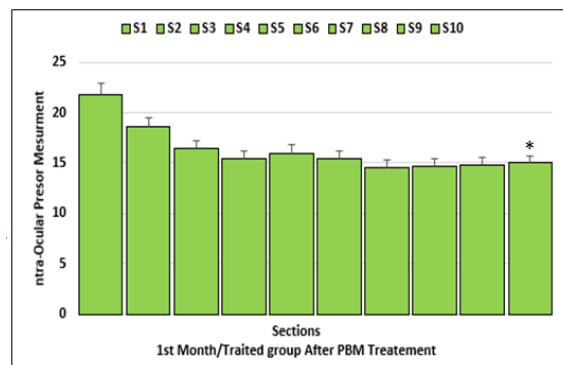
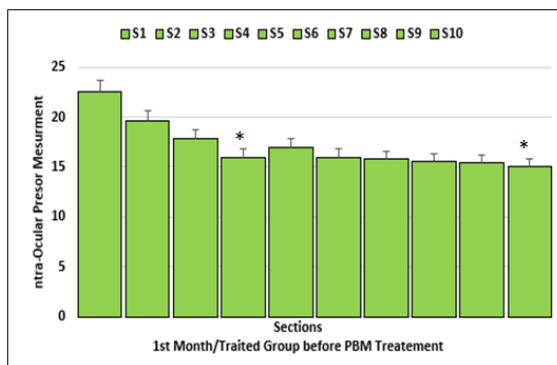
The statistical results were carried out with the EXCEL-Stat software, the average; the histograms and the P value ($p < 0.05$) were applied in order to highlight the results of our study.

3. RESULTS

20 patients aged (66 ± 8) (60% men and 40% women) were recruited in this study, divided into two groups (one group treated with PBM and the other considered as placebo group) with an IOP measurement before and after each treatment session.

The results obtained from IOP in summer marked and presented in the histograms below.

The results show a decrease in eye pressure, compared to the first session (from 22.6 mmHg before treatment to 15 mmHg after treatment session 10) during the first month.



* :significant
 ** :high significant
 * : $p < 0.05$

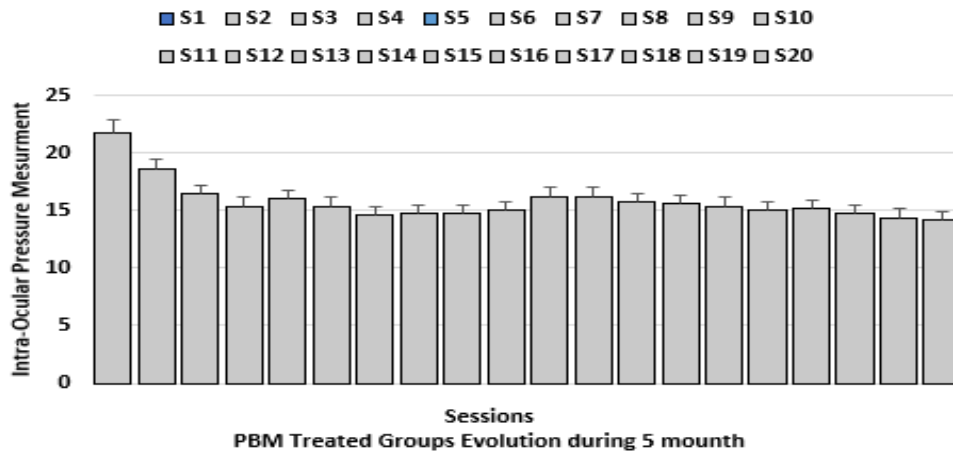


Fig. 3. Histograms showing the IOP evolution before and after treatment during 5 months

On the other hand, an insignificant decrease in the placebo group (from 14.8mmHg to 13.7 mmHg).

After 5 months the intraocular pressure was measured and the treatment was taking back the intraocular pressure content decreased significantly (from 16.6 mmHg to 14.2 mmHg) with an average of 2.2 mmHg.

The placebo group showed improvement (a slight decrease in intraocular pressure) from 16 mmHg to 14 mmHg with an average of 2 mmHg.

4. DISCUSSION

“Glaucoma is an optic neuropathy in which the primary risk factor is increased intraocular pressure (IOP), attributed to increased resistance to trabecular outflow of aqueous humor (AH). This resistance is believed to result from trabecular degeneration secondary to chronic oxidative stress and cellular senescence but may also involve inflammatory mechanisms whose roles are little known” [8].

“One of the major causes of the degeneration and cell loss of the glaucomatous trabecular meshwork would be the existence of chronic oxidative stress secondary to aging, and amplified in the event of glaucoma” [9]. “Indeed, in case of chronicity, chronic oxidative stress would promote senescence of trabecular tissue [10] cell apoptosis, accumulation of extracellular matrix (ECM) and stiffening of the cytoskeleton, leading to increased resistance to evacuation of the HA and thus to an elevation of the IOP” [9,11,12].

“The TM is the most sensitive tissue to oxidative damage in the anterior chamber” [13]. “Oxidative stress to the TM can cause much damage, such as reduce TM mitochondrial respiratory activity, leading to growth arrest [14], affect ECM structure [15] and lead to ECM accumulation [16], damage TM cellular DNA [17], alter membrane permeability [18], cause the rearrangement of TM cell cytoskeletal structures, cause the loss of cell-matrix adhesion [19], affect cell cycle progression [20], cause inflammatory cytokine release [21,22], and trigger apoptosis [23,24], as well as many forms of cell death” [25]. “Cell death may cause a free radical attack [26,27] and the loss or altered functionality of TM cells, leading to even more oxidative stress, thus beginning a vicious cycle” [28]. “At least, ROS alter the morphology, function and drainage of the anterior chamber filter channel that eventually leads to an increase in IOP” [29]. “In patients with glaucoma, the levels of mitochondrial DNA (mtDNA) damage and lipid peroxidation products in the TM are significantly higher compared with the controls [30,31] and their visual field defects, due to retinal ganglion cell degeneration, are directly proportional to oxidative damage to the TM” [32].

“An important principle of photobiomodulation therapy is that the dose of light administered does not exceed the damage threshold not be defined by their output power per session , but by the effective dose delivered to the target tissue at the specified wavelength” [33].

“Photobiomodulation (PBM) involves the use of red or near infrared light at low power densities to produce a beneficial effect on cells or tissues [34]. The main use proposed for

photobiomodulation therapy is to reduce inflammation [35] and to regenerate damaged tissues. The primary site of light absorption in mammalian cells has been identified as the mitochondria and, more specifically, cytochrome c oxidase (CCO)" [34].

"Some studies show that the purified enzyme, cytochrome c oxidase (CCO) was shown to be activated in vitro by red (633 nm) [36]. CCO is unit 4 of the mitochondrial respiratory chain and is a complex molecule with 13 separate protein subunits. CCO contains two different copper centers CuA and CuB and two heme centers, heme-a and heme-a3" [37].

"The leading hypothesis to explain how exactly light increases CCO enzyme activity is that nitric oxide (a molecule that is known to inhibit CCO by non-covalently binding between heme-a3 and CuB [38]) can be photodissociated by absorption of a photon of red or NIR light" [39]. to explain why PBM appears to have greater effects in diseased or damaged cells is that unhealthy or hypoxic cells are more likely to have inhibitory concentrations of NO, increasing the rate of respiration and ATP production. This proposed mechanism is

illustrated in (Fig. 4) which could explain the decrease in IOP after the session of PBM treatment.

"The significant decrease in IOP level after 5 months of PBM treatment could be explaining by that PBM may exert a prosurvival effect on cells via the activation of AKT/GSK3b/b-catenin pathway. Basically, protein kinase B (also known as AKT) can be activated by PBM, and then interact with glycogen synthase kinase 3b (GSK3b), inhibiting its activity. GSK3b is a serine-threonine kinase involved in cell death" [40]. "PBM activates Akt, which phosphorylates the Ser9 residue in GSK3b, rendering the enzyme inactive. b-catenin is an important component of Wnt signaling pathway but GSK3b-mediated phosphorylation of b-catenin or the tau protein seems to enhance TM cell death, and conversely phosphorylated GSK3b leads to cells survival" [41].

Another pathway that can be activated by PBM-mediated activation of Akt is "Akt/mTOR/cyclinD1, but It is not yet clear precisely how PBM activates Akt, and it is well known that Akt and ROS generation are closely intertwined" [42].

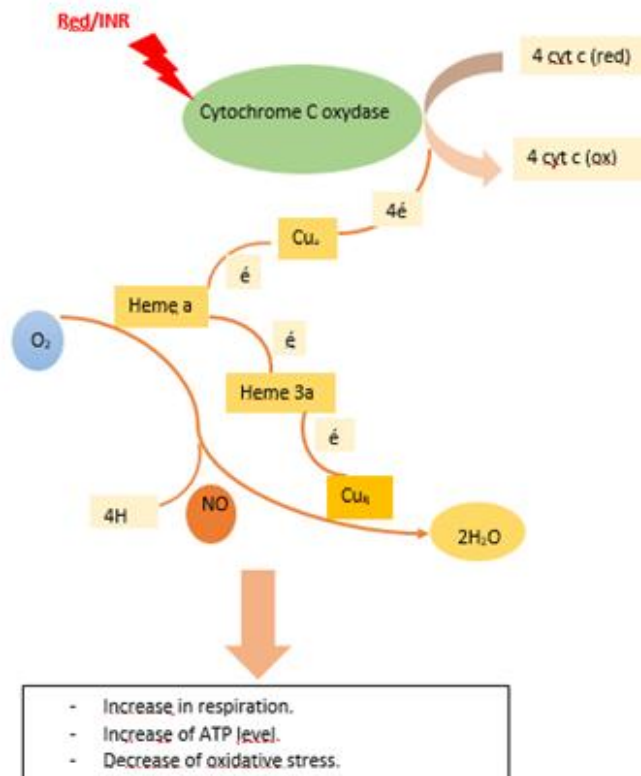


Fig. 4. The effect of RED/INR on the cytochrom C oxidase

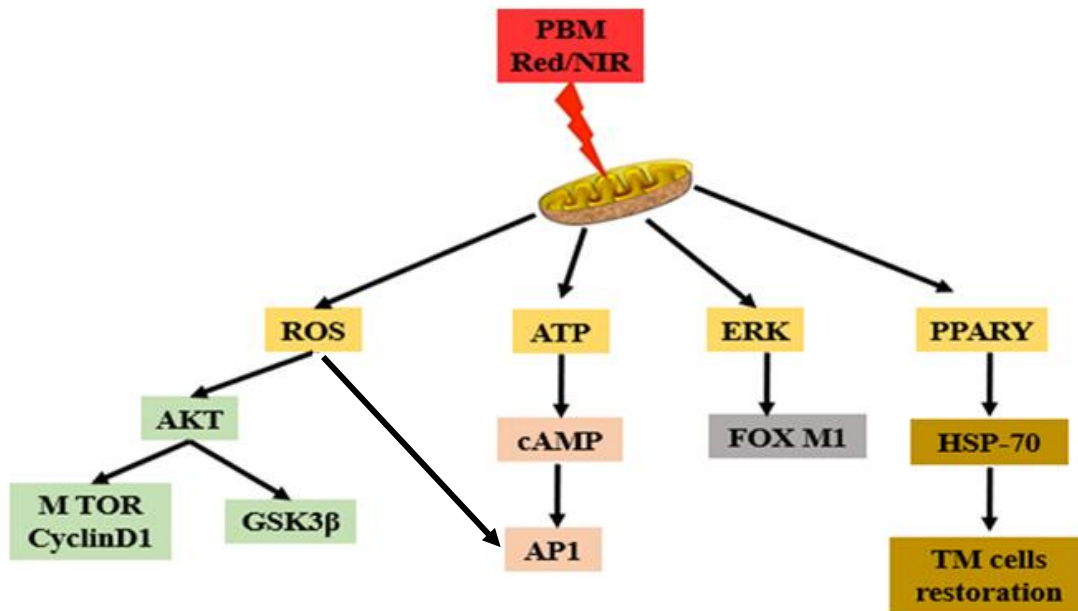


Fig. 5. Activation of transcription factors and signaling pathways after PBM

“Forkhead box protein M1 (FOXM1) is a transcription factor involved in the regulation of the transition from G1 to S phase of the cell cycle leading to mitotic division” [43] “FOXM1 is activated by epidermal growth factor via extracellular signal-regulated kinase (ERK)” [44]. “after PBM treatment, the mitogen-activated kinase (MEK)/ERK pathway was inhibited prevented the nuclear translocation of FOXM1, suggesting that Raf/MEK/MAPK/ERK signaling is crucial for the anticell senescence effect of PBM mediated by FOXM1” [45].

“Peroxisome proliferator-activated receptors (PPARs) play a role in the regulation of mitochondrial metabolism” [46] “They are nuclear receptors that regulate gene expression. PPAR-c is involved in the generation of heat shock protein 70 (HSP-70), which is anti-inflammatory [47] after PBM using INR in 850nm a marked rise in the expression of PPAR mRNA was observed, as well as increased PPAR-y activity, which decrease the inflammatory effect, by increasing the expression of a transcription factor that is signaling the synthesis of HSP70 and other anti-inflammatory proteins, leading to TM restoration” [48] (Fig. 5).

In fact, all these PBM pathways could promote anti-oxidant effect, inflammatory reduction, mitochondrial activity restoration and TM cells renewal, leading to a lower eye pressure and thus treat glaucoma.

5. CONCLUSION

In our study, photobiomodulation shows a clear improvement in the intraocular pressure compared to the placebo group, the use of this device in the daily practice of the ophthalmologist will improve the management of glaucomatous patients that could reduce the use of eye drops.

CONSENT

As per international standard or university standard, patient(s) written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

It is not applicable.

DISCLAIMER

Thera-RED light diode system was made by Laboratory Neurogenesis Research and Development patented by INAPI and certified by the national health organization (Algeria).

ACKNOWLEDGEMENT

We would like to thank all the authors; the clinical staff and the participants in this study.

COMPETING INTERESTS

Authors have declared that they have no known competing financial interests or non-financial

interests or personal relationships that could have appeared to influence the work reported in this paper.

REFERENCES

1. Rénard JP, Sellem E. Glaucoma primitif à angle ouvert. Rapport de la SFO. 2014;13-21.
2. Dana Blumberg, Alon Skaat, Jeffrey M. Liebmann. Emerging risk factors for glaucoma onset and progression; *Progress in Brain Research*. 2015;221:81-101. Available:<https://doi.org/10.1016/bs.pbr.2015.04.007>
3. Nan Zhang, Jiaying Wang, Ying Li, Bing Jiang. Prevalence of primary open angle glaucoma in the last 20 years: A meta-analysis and systematic review. *Nature portfolio Scientific Reports*. 2021;11:13762. Available:<https://doi.org/10.1038/s41598-021-92971>
4. Tham YC. et al. Global prevalence of glaucoma and projections of glaucoma burden through 2040: A systematic review and meta-analysis. *Ophthalmology*. 2014;121(11):2081–2090.
5. Colleen M McDowell, et al. Consensus recommendation for mouse models of ocular hypertension to study aqueous humor outflow and its mechanisms. *Investigative Ophthalmology & Visual Science*. 2022;63:12. DOI: 10.1167/iovs.63.2.12
6. Behnaz Ahrabi, Samareh Omidvari, Shamim Mollazadeh Ghomi, Navid Ahmady Roozbahany, Saeed Vafaei-Nezhad, Atefeh Shirazi Tehrani, Hojjat Allah Abbaszadeh, Shahram Darabi. Therapeutic effects of combination therapy and photobiomodulation therapy on retinal regeneration. *Journal of Lasers in Medical Sciences*. 2022;13:e36. DOI: 10.34172/jlms.2022.36
7. Cela D, Brignole-Baudouin F, Labbé A, Baudouin C. Glaucomatous trabeculum: An inflammatory trabeculopathy? *Journal Français d'Ophthalmologie*. 2022;45(4):455-477. Available:<https://doi.org/10.1016/j.jfo.2021.06.015>
8. Baudouin C, et al. Ocular surface inflammatory changes induced by topical antiglaucoma drugs: Human and animal studies. *Ophthalmology*; 1999.
9. Sabio G, et al. TNF and MAP kinase signalling pathways. *Semin Immunol*; 2014.
10. De La Luz Sierra M, et al. Differential processing of stromal-derived factor-1alpha and stromal-derived factor-1beta explains functional diversity. *Blood*; 2004.
11. Chen YM, et al. Tumor necrosis factor-alpha stimulates fractalkine production by mesangial cells and regulates monocyte transmigration: down-regulation by cAMP *Kidney Int*; 2003.
12. Izzotti A, Saccà SC, Longobardi M, Cartiglia C. Sensitivity of ocular anterior chamber tissues to oxidative damage and its relevance to the pathogenesis of glaucoma. *Invest Ophthalmol Vis Sci*. 2009;50:5251–5258.
13. Clopton DA, Saltman P. Low-level oxidative stress causes cell-cycle specific arrest in cultured cells. *Biochem Biophys Res Commun*. 1995;210:189–196.
14. Knepper PA, Goossens W, Palmberg PF. Glycosaminoglycan stratification of the juxtacanalicular tissue in normal and primary open-angle glaucoma. *Invest Ophthalmol Vis Sci*. 1996;37:2414–2425.
15. Knepper PA, Goossens W, Hvizd M, Palmberg PF. Glycosaminoglycans of the human trabecular meshwork in primary open-angle glaucoma. *Invest Ophthalmol Vis Sci*. 1996;37:1360–1367.
16. Izzotti A, Longobardi M, Cartiglia C, Saccà SC. Mitochondrial damage in the trabecular meshwork occurs only in primary open-angle glaucoma and in pseudoexfoliative glaucoma. *Plos One*; 6:e145672011.
17. Alvarado JA, Alvarado RG, Yeh RF, Franse-Carman L, Marcellino GR, Brownstein MJ. A new insight into the cellular regulation of aqueous outflow: How trabecular meshwork endothelial cells drive a mechanism that regulates the permeability of schlemm's canal endothelial cells. *Br J Ophthalmol*. 2005;89:1500–1505.
18. Zhou L, Li Y, Yue BY. Oxidative stress affects cytoskeletal structure and cell-matrix interactions in cells from an ocular tissue: The trabecular meshwork. *J Cell Physiol*. 1999;180:182–189.
19. Giancotti FG. Integrin signaling: Specificity and control of cell survival and cell cycle progression. *Curr Opin Cell Biol*. 1997;9:691–700.

20. Welge-Lüssen U, Birke K. Oxidative stress in the trabecular meshwork of POAG. *Klin Monbl Augenheilkd.* 2010;227:99–107.
21. Li G, Luna C, Liton PB, Navarro I, Epstein DL, Gonzalez P. Sustained stress response after oxidative stress in trabecular meshwork cells. *Mol Vis.* 2007;13:2282–2288.
22. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev.* 2002;82:47–95.
23. Frisch SM, Ruoslahti E. Integrins and anoikis. *Curr Opin Cell Biol.* 1997;9:701–706.
24. Martindale JL, Holbrook NJ. Cellular response to oxidative stress: Signaling for suicide and survival. *J Cell Physiol.* 2002;192:1–15.
25. Yan DB, Trope GE, Ethier CR, Menon IA, Wakeham A. Effects of hydrogen peroxide-induced oxidative damage on outflow facility and washout in pig eyes. *Invest Ophthalmol Vis Sci.* 1991;32:2515–2520.
26. Padgaonkar V, Giblin FJ, Leverenz V, Lin LR, Reddy VN. Studies of H₂O₂-induced effects on cultured bovine trabecular meshwork cells. *J Glaucoma.* 1994;3:123–131.
27. De La Paz MA, Epstein DL. Effect of age on superoxide dismutase activity of human trabecular meshwork. *Invest Ophthalmol Vis Sci.* 1996;37:1849–1853.
28. Wielgus AR, Sarna T. Ascorbate enhances photogeneration of hydrogen peroxide mediated by the iris melanin. *Photochem Photobiol.* 2008;84:683–691.
29. Kahn MG, Giblin FJ, Epstein DL. Glutathione in calf trabecular meshwork and its relation to aqueous humor outflow facility. *Invest Ophthalmol Vis Sci.* 1983;24:1283–1287.
30. Zanon-Moreno V, Marco-Ventura P, Lleo-Perez A, Pons-Vazquez S, Garcia-Medina JJ, Vinuesa-Silva I, Moreno-Nadal MA, Pinazo-Duran MD. Oxidative stress in primary open-angle glaucoma. *J Glaucoma.* 2008;17:263–268.
31. Jing Zhao, Shuang Wang, Wei Zhong, Ben Yang, Lixia Sun, Yajuan Zheng. Oxidative stress in the trabecular meshwork (review). *International Journal of Molecular Medicine.* 2016;38(4):995-1002. Available:<https://doi.org/10.3892/ijmm.2016.2714>
32. De Freitas LF, Hamblin MR. Proposed mechanisms of photobiomodulation or low-level light therapy. *IEEEJ Sel Top Quantum Electron.* 2016;22:348-364.
33. Michael R. Hamblin. Mechanisms and mitochondrial redox signaling in photobiomodulation. *Photochemistry and Photobiology.* 2018;94:199–212.
34. Peter Dungal, Joachim Hartinger, Sidrah Chaudary, Paul Slezak, Anna Hofmann, Thomas Hausner, Martin Strassl, Ernst Wintner, Heinz Redl, Rainer Mittermayr M. Low level light therapy by LED of different wavelength induces angiogenesis and improves ischemic wound healing. 2014;46(10):773-780. Available:<https://doi.org/10.1002/lsm.22299>
35. Pastore D, Greco M, Passarella S. Specific heliumneon laser sensitivity of the purified cytochrome c oxidase. *Int. J. Radiat. Biol.* 2000;76:863–870.
36. Karu TI, Kolyakov SF. Exact action spectra for cellular responses relevant to phototherapy. *Photomed. Laser Surg.* 2005;23:355–361.
37. Sarti P, Forte E, Mastronicola D, Giuffre A, Arese M. Cytochrome c oxidase and nitric oxide in action: Molecular mechanisms and pathophysiological implications. *Biochim. Biophys. Acta.* 2012;1817:610–619.
38. Lane N. Cell biology: Power games. *Nature.* 2006;443:901–903.
39. Ma T. GSK3 in Alzheimer's disease: Mind the isoforms. *J. Alzheimers Dis.* 2014;39:707–710.
40. Liang J, Liu L, Xing D. Photobiomodulation by lowpower laser irradiation attenuates Abeta-induced cell apoptosis through the Akt/GSK3beta/beta-catenin pathway. *Free Radic. Biol. Med.* 2012;53:1459–1467.
41. Dolado I, Nebreda AR. AKT and oxidative stress team up to kill cancer cells. *Cancer Cell.* 2008;14:427–429.
42. Halasi M, Gartel AL. Targeting FOXM1 in cancer. *Biochem. Pharmacol.* 2013;85:644–652.
43. Xie Y, Cui D, Sui L, Xu Y, Zhang N, Ma Y, Li Y, Kong Y. Induction of forkhead box M1 (FoxM1) by EGF through ERK signaling pathway promotes trophoblast cell invasion. *Cell Tissue Res.* 2015;362:421–430.
44. Eells JT, Henry MM, Summerfelt P, Wong-Riley MT, Buchmann EV, Kane M, Whelan NT, Whelan HT. Therapeutic photobiomodulation for methanol-induced retinal toxicity. *Proc. Natl Acad. Sci. USA.* 2003;100:3439–3444.

45. Mello T, Materozzi M, Galli A. PPARs and mitochondrial metabolism: From NAFLD to HCC. PPAR Res. 2016;7403230.
46. Croasdell A, Duffney PF, Kim N, Lacy SH, Sime PJ, Phipps RP. PPARgamma and the innate immune system mediate the resolution of inflammation. PPAR Res. 2015;549691.
47. de Lima FM, Albertini R, Dantas Y, Maia-Filho AL, Santana Cde L, Castro-Faria-Neto HC, Franca C, Villaverde AB, Aimbire F. Low-level laser therapy restores the oxidative stress balance in acute lung injury induced by gut ischemia and reperfusion. Photochem. Photobiol. 2013; 89:179–188.
48. Tang D, Kang R, Xiao W, et al. The anti-inflammatory effects of heat shock protein 72 involve inhibition of high-mobility-group box 1 release and proinflammatory function in macrophages. The Journal of Immunology. 2007;179(2):1236-44.

© 2023 Boudghene et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/98246>