



International Journal of Biochemistry Research & Review 12(3): 1-10, 2016, Article no.IJBCRR.25627 ISSN: 2231-086X, NLM ID: 101654445

> SCIENCEDOMAIN international www.sciencedomain.org

## Sulphasalazine Prevents Fibrosis; Relevance of TGFβRI

## Elsayed Gomaa Elsayed Elsakka<sup>1\*</sup>, Gamil Mohammed Abd-Allah<sup>1</sup> Ahmed Ibrahim Abulsoud<sup>1</sup>, Ahmed Mohammed Ibrahim Mansour<sup>2</sup> and Saved Abdel Raheem<sup>3</sup>

<sup>1</sup>Department of Biochemistry, Faculty of Pharmacy, Alazhar University, Cairo, Egypt. <sup>2</sup>Department of Pharmacology and Toxicology, Faculty of Pharmacy, Alazhar University, Cairo, Egypt. <sup>3</sup>Department of Pathology, Faculty of Medicine, Alazhar University, Cairo, Egypt.

## Authors' contributions

This work was carried out in collaboration between all authors. Authors EGEE, GMAA and AMIM designed the study, wrote the protocol and supervised the work. Authors EGEE and AMIM carried out the animal modeling. Authors EGEE and AIA carried out laboratory work, performed the statistical analysis, wrote the first draft of the manuscript, managed the literature searches and edited the manuscript. Author SAR carried out the histopathological assessment. All authors read and approved the final manuscript.

#### Article Information

DOI: 10.9734/IJBCRR/2016/25627 Editor(s): (1) Hector M. Mora Montes, Department of Biology, University of Guanajuato, Mexico. (2) Richard A. Manderville, Departments of Chemistry and Toxicology University of Guelph, Canada. Reviewers: (1) Anonymous, China Three Gorges University, China. (2) Sahar S. Abd El-Rahman, Cairo University, Giza, Egypt. (3) Anonymous, Sapienza University, Rome, Italy. (4) Sura Wanessa Santos Rocha, Research Center Aggeu Magalhaes, Brazil. (5) Darko Nozic, Military Medical Academy, Belgrade, Serbia. Complete Peer review History: http://sciencedomain.org/review-history/14979

**Original Research Article** 

Received 12<sup>th</sup> March 2016 Accepted 3<sup>rd</sup> June 2016 Published 10<sup>th</sup> June 2016

## ABSTRACT

Aims: The aim of this study is to investigate the protective effect of sulphasalazine on liver fibrosis. Besides; investigation of the expression pattern of transforming growth factor  $\beta$  receptor I (TGF $\beta$ RI) in liver fibrosis and the possible modulatory effect of sulphasalazine on it. Study Design: Controlled experiment.

Place and Duration of Study: Department of Biochemistry and Department of Pharmacology and Toxicology, Faculty of pharmacy (boys) Al-Azhar University, between February 2015 and June 2015.

\*Corresponding author: E-mail: elsayedelsakka750@yahoo.com, elsayedelsakka750@azhar.edu.eg;

**Methodology:** Five Sprague-Dawley rats groups were used for the experiment. Control group: Receiving corn oil; DMSO group: Injected with DMSO; 6 weeks group: Injected with 50% CCl<sub>4</sub> in corn oil; sulphasalazine group: Injected with sulphasalazine and CCl<sub>4</sub> and finally mesalazine group: were given mesalazine and CCl<sub>4</sub>. On the day after the last dose, rats were anesthetized with diethyl ether and blood samples were collected for measurement of blood chemistry. The animals then were euthanized, and livers were harvested and divided into 2 parts; one part was processed for standard histology and immunofluorescence techniques and the other was homogenized for oxidative status assessment.

**Results:** TGF $\beta$ RI has been shown to be upregulated in chronic liver injury; fibrosis stage; with expression occurring mainly in cell membrane of lesion area. This expression was decreased significantly with sulphasalazine treatment. Sulphasalazine has shown to have ability to diminish fibrosis in chronic liver injury. This decrease in fibrosis observed with sulphasalazine was in parallel with decrease in TGF $\beta$ RI expression. Besides; this action of sulphasalazine has been suggested to be due to the whole molecule not to its moiety; mesalazine.

**Conclusion:** TGF $\beta$ RI may be used as a candidate marker for diagnosis and prognosis of chronic liver diseases. Besides; it may be used as a target therapy for chronic liver diseases. Moreover; sulphasalazine might be a promising adjuvant therapy for chronic liver diseases.

*Keywords:* TGFβRI; fibrosis; sulphasalazine; mesalazine.

#### **1. INTRODUCTION**

Liver plays an unlimited role in the preservation and body homeostasis regulation. It is included in most biochemical pathways to growth, protection against disease, nutrient fund, energy facility and reproduction [1].Hepatotoxicity can be generated by certain common causes including therapeutic agents, natural chemicals, laboratory and manufacturing agents and herbal therapies [2].

Transforming growth factor  $\beta$  receptors (TGF $\beta$ Rs) are of 2 types TGF- $\beta$ RI and TGF- $\beta$ RI. They mediate Hepatic stellate cells (HSCs) activation [3] after binding to their ligand cytokine TGF $\beta$ , the profibrogenic cytokine that is traditionally considered the key fibrogenic stimulus to HSC [4].

Carbon tetra chloride (CCl<sub>4</sub>) is one of the most communal models for inducing hepatotoxicity. It is transformed into a toxic CCl<sub>3</sub><sup>-</sup> radical by hepatic cytochrome P 450 2E1 (CYP2E1). Thus; it brings an acute Centro-lobular necrosis which starts a wound healing response (fibrosis) [5]. However, Contact to these chemicals in humans is rare and generally occurs in the manufacturing during fabrication and in places where these chemicals is usually used [6].

Hepatic stellate cells are the most important player in hepatic injury. Quiescent HSCs are characterized by significant desmin expression and vitamin A storage. Following liver injury, HSCs lose their vitamin A content, rise the expression of  $\alpha$ -smooth muscle actin ( $\alpha$ -SMA) and gain a myofibroblast- like phenotype [7]. These events are primarily triggered by mediators released by activated Kupffer cell and injured hepatocytes [8].

Sulfasalazine is a synthetic drug obtained from the grouping of sulfapyridine and 5-aminosalicylic acid (mesalazine), an antibiotic and an antiinflammatory agent, respectively. This drug is usually used in the inflammatory diseases of the large intestine and rheumatoid arthritis [9]

Despite great knowledge about liver fibrosis, there are little drugs approved for management of it. So the aim of this work is to investigate the possible protective effect of sulphasalazine on liver fibrosis; an aspect that has gone side by side with investigation of expression pattern of TGF $\beta$ RI in liver fibrosis and investigation of the possible modulatory effect of sulphasalazine on it.

## 2. METHODOLOGY

#### 2.1 Animal Model

Adult male Sprague-Dawley (SD) rats weighing 250–300 g and aging 70 days were used in the current study. The animals were obtained from the breeding colony maintained at the animal house of the Nile Company for pharmaceuticals, Cairo, Egypt. They were housed in the animal facility of Faculty of Pharmacy, Al-Azhar University in 20 X 18 X 25 cm plastic cages with stainless steel wire lids and mesh floor with 5 animals per cage. They were kept at  $23\pm1^{\circ}$ C, 55% relative humidity with 12:12-h light: Dark cycle. They were maintained on a standard

rodent chow (El-Nasr Company, Abou-Zaabal, Cairo, Egypt) and allowed to food and water ad libitum. Animals were randomly divided into five groups; (1) control group: 9 rats received corn oil (2 mg/kg/twiceweekly/intraperitoneal(IP)/ repeated for 6 weeks); (2)DMSO group: 12 rats were injected with DMSO, the vehicle of sulphasalazine (1 ml/kg/day/IP/repeated for 6 weeks) and 50% CCl₄ in corn oil (4 ml/kg/IP/twice weekly/repeated for 6 weeks); (3) 6 weeks group:12 rats were injected with 50% CCl<sub>4</sub> in corn oil (4 ml/kg/IP/twice weekly/repeated for 6 weeks); (4) sulphasalazine group: 12 rats injected with sulphasalazine were (75 mg/kg/day/IP/repeated for 6 weeks) [10] and CCl<sub>4</sub> as previously indicated in 6 weeks group and (5) mesalazine group: 12 rats were given mesalazine (100 mg/kg/day/orally by oral gavage/repeated for 6 weeks) [11] and CCl<sub>4</sub> as previously indicated in 6 weeks group. At the of experimental period, rats end were anesthetized with diethyl ether and blood samples have been drawn from retro orbital plexus for measurement of blood chemistry. The animals then were euthanized, and tissue samples from the livers were harvested and divided into 2 parts; one part was processed by standard histology and immunofluorescence techniques and the other was homogenized in 0.15 M KCl for oxidative stress assay. All animal procedures were performed in accordance with the international guide for the care and use of laboratory animals [12].

## 2.2 Antibodies and Chemicals

Rabbit polyclonal TGFβR1 antibody was purchased from Santa Cruz Biotechnology (CA, USA). Cy3-conjugated goat anti-rabbit antibody was purchased from Jackson Immunoresearch (PA, USA). 4, 6- Diamidino-2- phenyl indole (DAPI). CCl<sub>4</sub> and DMSO were purchased from Sigma-Aldrich (MO, USA). Sulphasalazine was kindly supplied as yellowish powder by El-Kahera Pharmaceuticals Company, Cairo, Egypt and mesalazine was kindly supplied as white powder by Pharopharm pharmaceuticals company, Alexandria, Egypt.

## 2.3 Biochemical Analysis

Serum enzymatic activities of transaminases (ALT and AST) were estimated by kinetic method according to the method of international federation of clinical chemistry (IFCC) [13,14]. Alkaline phosphatase (ALP) activity was assayed according to the method of IFCC [15-17] and

serum albumin concentration according to the method described by Gendler [18]. The liver homogenate was used for the determination of the oxidative stress parameters. The level of thiobarbituric acid-reactive substances was assayed as malondialdehyde (MDA) as described by Mihara and Uchiyama [19]. The activity of SOD was determined using the method described by Marklund [20].

## 2.4 Immunofluorescence Analysis

Liver tissues sections were handled according to method described by Abdel-Bakky et al. [21]. The primary TGF $\beta$ R1 antibody was diluted in blocking solution in the suitable dilution (1:400) and left overnight in 4°C. Secondary antibody (cyanine red conjugated) diluted in the blocking solution was incubated for 30 min and the nuclei were counterstained using DAPI. Finally, all slides were mounted with the fluoromount solution, covered by covering slips, and allowed to stand for detection by immunofluorescence microscope (Leica DM 5500B).

## 2.5 Histopathological Examination

Liver samples were taken from rats belonging to the different groups and fixed in 10% neutral buffered formalin for 24 hours. Washing was done in tap water, and then serial dilutions of alcohol (methyl, ethyl and absolute ethyl) were used for dehydration. Specimens were cleared in xylene and embedded in paraffin at 56°C in hot air oven for 24 h. Paraffin wax tissue blocks were prepared for sectioning at 4 microns thickness by sledge microtome.

The obtained tissue sections were collected on glass slides, deparaffinized, stained by hematoxylin & eosin stain and then examination was done through a light electric microscope [22]. Histopathological grading was achieved by subjective scoring approved by pathology department, faculty of pharmacy, Al-Azhar university, Cairo, Egypt with assistance of attached clues.

## 2.6 Statistical Analysis

Data were presented as the mean ±SE. Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by Tukey-Kramer as a post test, according to the number. of groups. The 0.05 level of probability was used as the criterion of significance using GraphPad Prism software version 5 (GraphPad Software Inc, CA, USA).

## 3. RESULTS

## 3.1 Liver Functions and Oxidative Status Assessment

To assess liver functions; we have performed liver function tests including serum transaminases, ALP, and albumin. The oxidative status of liver tissues has been determined through MDA and SOD assay as indicated in Table 1 and Table 2.

As indicated in Table 1 and Table 2; Serum ALT and AST activity of control group rats were  $46.6\pm2.85$  and  $54.1\pm2.65$  respectively. Administration of CCl<sub>4</sub> I.P. alone or combined with DMSO for 6 weeks significantly increased ALT and AST activity compared to control group. Interestingly concomitant administration of CCl<sub>4</sub> with sulphasalazine reduced serum activity of AST compared to DMSO group.

As shown in Table 1 and Table 2; ALP activity of control group rats was  $312\pm21.9$ . Administration of CCl<sub>4</sub> I.P. for 6 weeks increased ALP significantly by 83% if given only and by 86% in combination with DMSO compared to control group. Concomitant administration of sulphasalazine with CCl<sub>4</sub> decreased ALP activity by 10% compared to DMSO group.

Serum albumin of control group was  $3.81\pm0.08$  g/dl. A significant decrease was observed only in mesalazine group as shown in Table 1 and Table 2.

As shown in Table 1 and Table 2; the hepatic MDA content of control group was  $4.73\pm0.318$  nmol/g tissue. This content was significantly increased on CCl<sub>4</sub> treated groups to be  $31.9\pm1.44$  (nmol/g tissue) for 6 weeks group,  $31.7\pm1.82$  (nmol/g tissue) for DMSO group and  $25.7\pm1.5$  (nmol/g tissue) for sulphasalazine group. On the other hand; combination of CCl<sub>4</sub> with mesalazine showed MDA content of  $9.86\pm0.339$  respectively; results that showed significant decrease compared to 6 weeks CCl<sub>4</sub> treatment although they appeared non-significantly changed from control group. Finally sulphasalazine showed a significant decrease in MDA content compared to DMSO group.

Table 1 and Table 2 show the M  $\pm$  SEM for SOD of studied groups. The control group showed tissue SOD activity of 526 $\pm$ 25.2 U/mg tissue. This activity was significantly decreased in other groups to become 242 $\pm$ 15.4, 223 $\pm$ 14, 281 $\pm$ 16 and 225 $\pm$ 9.65 for DMSO, 6 weeks, sulphasalazine, and mesalazine groups.

#### 3.2 Expression and Localization of TGFBR1

Fig. 1 and Graph 1 show that TGF $\beta$ R1 protein showed minimal expression in normal liver tissues. The tissue expression was showed to be maximal in 6 weeks CCl<sub>4</sub> groups. It is noted that the expression increased by about 146% in 6 weeks CCl<sub>4</sub> treatment group. It is also observed that the expression was located in the cell membrane of epithelial hepatic tissue in areas that shows great lesions.

 Table 1. Liver functions and oxidative status assessment tests (samples were taken from all animals belonging to each group)

	ALT (IU/L)	AST (IU/L)	ALP (IU/L)	Albumin (g/dl)	MDA (nmol/g tissue)	SOD (U/mg tissue)
Control CCl <sub>4</sub> 6 weeks	46.6±2.85 349±13.4 <sup>(a)</sup>	54.1±2.65 436±15.4 <sup>(a)</sup>	312±21.9 570±24.5 <sup>(a)</sup>	3.81±0.08 3.39±0.13	4.73±0.32 31.9±1.44 <sup>(a)</sup>	526±25.2 223±14 <sup>(a)</sup>
CCl₄ 6 weeks I.P. +DMSO I.P.	332±12.8 <sup>(a)</sup>	402±8.91 <sup>(a)</sup>	580±21.1 <sup>(a)</sup>	3.35±0.12	31.7±1.82 <sup>(a)</sup>	242±15.4 <sup>(a)</sup>
CCl₄ 6 weeks I.P. + sulphasalazine I.P.	328±19 <sup>(a)</sup>	338±14.5 <sup>(a)(b)(c)</sup>	523±19.8 <sup>(a)</sup>	3.34±0.12	25.7±1.5 <sup>(a)</sup>	281±16 <sup>(a)</sup>
CCl <sub>4</sub> 6 weeks I.P. + oral mesalazine	315±19 <sup>(a)</sup>	486±24.2 <sup>(a)</sup>	701±44 <sup>(a)(b)</sup>	3.18±0.16 <sup>(a)</sup>	9.86 ±0.34 <sup>(b)</sup>	225±9.65 <sup>(a)</sup>

Data are expressed as mean  $\pm$  SEM

(a) Significantly different from control group

(b) Significantly different from 6 weeks CCl<sub>4</sub> group

(c) Significantly different from 6 weeks CCl₄ + DMSO (for sulphasalazine) using one-way ANOVA followed by Tukey-Kramer test for multiple comparison test at P≤0.05

	ALT (%change)	AST (%change)	ALP (%change)	Albumin (%change)	MDA (%change)	SOD (%change)
Control CCl <sub>4</sub> 6 weeks I.P.	100±6.12 749±28.8 <sup>(a)</sup>	100±4.9 806±28.47 <sup>(a)</sup>	100±7.02 183±7.85 <sup>(a)</sup>	100±2.1 89±3.41	100±6.77 674±30.4 <sup>(a)</sup>	100±4.79 42.4±2.66 <sup>(a)</sup>
CCl₄ 6 weeks I.P. +DMSO I.P.	712±27.5 <sup>(a)</sup>	743±16.5 <sup>(a)</sup>	186±6.76 <sup>(a)</sup>	87.9±3.15	670±38.5 <sup>(a)</sup>	46±2.93 <sup>(a)</sup>
CCl <sub>4</sub> 6 weeks I.P. + sulphasalazine	704±40.8 <sup>(a)</sup>	625±26.8 <sup>(a)(b)(c)</sup>	168±6.35 <sup>(a)</sup>	87.7±3.15	543±31.7 <sup>(a)</sup>	53.4±3.04 <sup>(a)</sup>
I.P. + oral mesalazine	676±40.8 <sup>(a)</sup>	898±44.7 <sup>(a)</sup>	225±4.1 <sup>(a)(b)</sup>	83.5±4.2 <sup>(a)</sup>	208 ±7.19 <sup>(b)</sup>	42.8±1.83 <sup>(a)</sup>

 Table 2. Liver functions and oxidative status assessment tests (percentage change from control) (samples were taken from all animals belonging to each group)

	DAPI	TGFβR1	Merge
control	<u>0 μm 100</u>	C.V.	
6 weeks I.P. CCL <sub>4</sub>	<u>0 μm 100</u>		
6 weeks I.P. CCL <sub>4</sub> + DMSO	<u>0 μm 100</u>		
6 weeks I.P.CCL <sub>4</sub> + sulphasalazine	0 µm 100		
6 weeks I.P.CCL <sub>4</sub> + mesalazine	<u>0 µm 100</u>		

Fig. 1. Immunofluorescence staining of liver sections of studied groups showing minimal TGFβR1 expression in control group. Six weeks CCl<sub>4</sub> treated group shows maximal expression located mainly in cell membrane (yellow rectangle). It is noted also that there is non-significant expression change between 6 weeks CCl<sub>4</sub> treatment alone or combined with DMSO. Combination of CCl<sub>4</sub> with sulphasalazine or mesalazine show decrease in the expression compared to 6 weeks CCl<sub>4</sub> treatment with maximal decrease occurring with sulphasalazine

It is also noted that concomitant administration of DMSO and  $CCl_4$  for 6 weeks produced nonsignificant change from  $CCl_4$  only. On the other hand; Concomitant administration of sulphasalazine and  $CCl_4$  decreased the expression by about 46% compared to DMSO group while concomitant administration of mesalazine with  $CCl_4$  reduced the expression by about 36% compared to 6 weeks  $CCl_4$  group.



Graph 1. Fluorescence intensity was obtained from 5 fields of each liver section (minimally 2 rats of each group) using ImageJ software Data are expressed as mean ± SEM

 (a) Significantly different from control group
 (b) Significantly different from 6 weeks CCl₄ group
 (c) Significantly different from 6 weeks CCl₄ + DMSO
 using one-way ANOVA followed by Tukey-Kramer test for multiple comparison test at P≤0.05

#### 3.3 Histopathological Findings

Effects of CCl<sub>4</sub> treatment on histopathological findings of liver tissue are represented in Table 3 and Fig. 2; the control group shows normal hepatic architecture. On contrast, six weeks CCl<sub>4</sub> treatment shows expanded portal tract with fibrous septa extending from a portal tract to another (yellow arrows), marked micro- and macro-vesicular steatosis. The hepatocytes exhibit some aspects of single cell necrosis with dilated central veins and mild interface activity.

It is noted that sulphasalazine had minimal effect on steatosis. Interestingly; despite sulphasalazine group animals show marked steatosis, they also show completely diminished underlying fibrosis in their liver tissues. Finally; combination of mesalazine with CCl<sub>4</sub> results in some alleviation in injury signs. However; it seems not to have positive effects regarding steatosis or fibrosis.

Histopathological micrograph of liver samples of CCl<sub>4</sub> treated groups combined with DMSO, sulphasalazine and mesalazine x235 using H and E stain. Control: liver tissue showing average PT (yellow arrows), average central vein (red arrow), and average hepatocytes arranged in cords. 6 weeks I.P. CCl<sub>4</sub>: Liver tissue showing expanded PT with underlying fibrosis and fibrous septa extending from PT to PT (yellow arrows), marked micro- and macro-vesicular steatosis. 6 weeks I.P. CCl<sub>4</sub> + I.P. DMSO: Liver tissue showing average portal tract (yellow arrows), fibrous septa extending from PT (blue arrows), average hepatocytes and moderate steatosis. 6 weeks I.P. CCl<sub>4</sub> + I.P. sulphasalazine: Liver tissue showing average PT (yellow arrow), and marked steatosis. 6 weeks I.P. CCl<sub>4</sub> + oral mesalazine: Liver tissue showing average PT (yellow arrow), short fibrous septa (red arrow), average hepatocytes and moderate steatosis.

#### 4. DISCUSSION

Administration of I.P. CCl<sub>4</sub> for 6 weeks has significantly elevated serum transaminases enzymes activities suggesting hepatocellular damage. These results agreed with previous reports that CCl<sub>4</sub> significantly increases serum transaminases [23-25]. This CCl<sub>4</sub> induced hepatic damage has been reported to be due to oxidative stress [26,27]. Concomitant administration of sulphasalazine with CCl<sub>4</sub> has reduced AST not ALT activity compared to DMSO group suggesting the positive effect toward liver injury. These effects might be due to the anti-inflammatory action of sulphasalazine. This disagrees with what has been reported by Jennings and his coworkers a result that was interpreted by the relationship of hepatic injury, steatosis, fibrosis and cirrhosis to ulcerative colitis not sulphasalazine medication [28].

ALP activity caused by IP CCl<sub>4</sub> administration for 6 weeks and DMSO group has showed significant increase compared to control which was less than 3 times as control; a result that might suggest hepatotoxicity and mild biliary toxicity. This matches with that previously reported by Posen and Doherty [29]. Concomitant administration of mesalazine and CCl<sub>4</sub> for 6 weeks has significantly elevated ALP activity compared to 6 weeks CCl<sub>4</sub> treatment. This result might suggest the toxic effect of mesalazine with the mentioned regimen on hepatic tissue or synergistic effect of mesalazine on CCl<sub>4</sub> toxicity; a suggestion that might be clear upon ALP and albumin result.

Meanwhile; it was clarified that free radicals production and oxidative stress are main players in liver injury especially  $CCl_4$  induced liver injury [26,27]. In our study; administration of  $CCl_4$  has significantly elevated tissue content of MDA and reduced serum SOD activity. These results

agreed with the previous reports that demonstrated the great role of oxidative stress in  $CCl_4$  induced liver injury [24,30,31].

Supporting the measured biochemical data: Our histopathological findings have demonstrated that administration of  $CCl_4$  for 6 weeks produced fibrosis of peri-portal or portal-portal fibrosis with intact architecture. This is similar to what has been reported that continued administration of  $CCl_4$  leads to hepatic fibrosis, cirrhosis, and hepatocellular carcinoma [32].

# Table 3. Histopathological findings of the studied groups (minimally 2 rats of each group with5 fields of each rat at least)

	Control	CCl₄ 6 weeks	CCl₄ 6 weeks + DMSO	CCl <sub>4</sub> 6 weeks + sulphasalazine	CCl <sub>4</sub> 6 weeks + mesalazine
CV	0	+	0	0	0
Steatosis	0	+++	++	+++	++
Hepatocyte	0	+	0	0	0
Spotty necrosis	0	0	+	0	0
Interface activity	0	+	0	0	0
PT	0	+	++	0	0
Fibrosis	0	++	++	0	++

Central vein (CV): 0: within normal +: dilated ++: markedly dilated

Steatosis, Spotty necrosis, Interface activity: 0: no +: mild ++: moderate to marked Hepatocytes: 0: within normal +: single cell necrosis++: confluent or diffuse necrosis Portal tract: 0: within normal +: expanded ++: expanded with inflammatory infiltrate

Fibrosis: 0: no fibrosis

+: fibrosis confined to enlarged portal zones

++: fibrosis of peri-portal or portal-portal septa with intact architecture

+++: architectural distortion (septal or bridging fibrosis) without obvious cirrhosis

++++: probable or definite cirrhosis



Fig. 2. Effect of CCl₄ administration on hepatic histopathological findings with investigation of the possible modulatory effect of sulphasalazine and mesalazine (minimally 2 rats of each group with 5 fields of each rat at least)

It has been found that concomitant administration of sulphasalazine and CCl<sub>4</sub> had markedly cleared fibrosis; major findings seen in CCl<sub>4</sub>. However, there is no positive effect on steatosis. This may be an expected result according to in-vitro study reports of Oakley et al. [33] concerning the inhibitory effect of sulphasalazine on the machinery and machinery of fibrous tissue synthesis.

As an interesting matter; the decrease in histopathological scores observed in mesalazine group didn't agree with sulphasalazine group although it was documented that sulphasalazine is a prodrug for mesalazine in some pharmacological actions [34]. This might suggest that the action of sulphasalazine previously reported by Oakley et al. [33] may be related to the sulphasalazine molecule at all not related to its moiety, mesalazine.

The TGF-β is the principal regulator in chronic liver injury sharing in all stages of disease progression [35]. Its action begins by binding to its receptor TGFBRI and TGFBRII [36]. It has been observed that TGFBR1expression occurred in the cell membrane. This agrees with Massague and Chen reports [37]. It has been also shown that TGFBR1 expression was upregulated only in chronic liver toxicity; specifically phases that showed some extent of tissue remodeling prescribed by Devaux et al. [38] suggesting involvement of TGFBR1in cellular processes involved in chronic not acute liver injury. This was similar to the previous reports in myocardial infarction (MI) by Devaux et al. [38]. Besides; they also showed that the maximal expression occurs in lesion areas suggesting the direct relationship between TGFBR1expression and lesions grade; a result that also is guite similar to Devaux et al. [38]. It was also observed that TGFBR1expression was nearly diminished in sulphasalazine group suggesting the tight linkage between TGFBR1 expression and fibrosis but not steatosis. This result can be explained by pro apoptotic effect of sulphasalazine toward activated HSC, the major cellular promoter of fibrogenesis [39].

## 5. CONCLUSION

Our findings indicate, for the first time, that TGF $\beta$ R1 content is increased in chronic liver injury. TGF $\beta$ R1 is upregulated in injury combined with underlying fibrosis; a result that may suggest usage of TGF $\beta$ R1 as a candidate marker for diagnosis and prognosis of chronic liver diseases

and a target for liver disease therapy. Moreover, sulphasalazine exhibits a fibroprotective effect in experimental liver fibrosis. This suggests a possible use of sulphasalazine as adjuvant in therapy of chronic liver diseases.

## ETHICAL APPROVAL

All authors hereby declare that all experiments have been examined and approved by the appropriate ethics committee of Alazhar University.

## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## REFERENCES

- Sharma A, Chakraborti KK, Handa SS. Antihepatotoxic activity of some Indian herbal formulations as compared to silymarin. Fitoterapia. 1991;62:229-35.
- Ostapowicz G, Fontana RJ, Schiodt FV, Larson A, Davern TJ, Han SH, et al. Results of a prospective study of acute liver failure at 17 tertiary care centers in the United States. Ann Intern Med. 2002;137(12):947-54.
- Roberts AB. Tgf-beta signaling from receptors to the nucleus. Microbes Infect. 1999;1(15):1265-73.
- Tsukamoto H. Cytokine regulation of hepatic stellate cells in liver fibrosis. Alcohol Clin Exp Res. 1999;23(5):911-6.
- Domenicali M, Caraceni P, Giannone F, Baldassarre M, Lucchetti G, Quarta C, et al. A novel model of CCl<sub>4</sub>-induced cirrhosis with ascites in the mouse. J Hepatol. 2009;51(6):991-9.
- 6. Mormone E, George J, Nieto N. Molecular pathogenesis of hepatic fibrosis and current therapeutic approaches. Chem Biol Interact. 2011;193(3):225-31.
- Gressner AM. Transdifferentiation of hepatic stellate cells (ITO cells) to myofibroblasts: A key event in hepatic fibrogenesis. Kidney Int Suppl. 1996;54:s39-s45.
- Nieto N. Oxidative-stress and il-6 mediate the fibrogenic effects of [corrected] kupffer cells on stellate cells. Hepatology. 2006;44(6):1487-501.
- 9. Ardizzone S, Bianchi PG. A practical guide to the management of distal ulcerative colitis. Drugs. 1998;55(4):519-42.

- Tugcu V, Ozbek E, Tasci AI, Kemahli E, Somay A, Bas M, et al. Selective nuclear factor kappa-b inhibitors, pyrolidium dithiocarbamate and sulfasalazine, prevent the nephrotoxicity induced by gentamicin. Bju Int. 2006;98(3):680-6.
- Hayashi Y, Aoyagi K, Morita I, Yamamoto C, Sakisaka S. Oral administration of mesalazine protects against mucosal injury and permeation in dextran sulfate sodiuminduced colitis in rats. Scand J Gastroenterol. 2009;44(11):1323-31.
- Institute of Laboratory Animal Resources (US). Use of Laboratory Animals, National Institutes of Health (US). Guide for the care and use of laboratory animals. National Academies; 1985.
- 13. Bergmeyer HU, Horder M, Rej R. International Federation of Clinical Chemistry (IFCC) Scientific Committee, analytical section: Approved recommendation (1985) on IFCC methods for the measurement of catalvtic concentration of enzymes. Part 2. IFCC method for aspartate aminotransferase (Iaspartate: 2-oxoglutarate aminotransferase, ec 2.6.1.1). J Clin Chem Clin Biochem. 1986;24(7):497-510.
- 14. Bergmeyer HU, Horder M, Rej R. International Federation of Clinical Chemistry (IFCC) Scientific Committee, analytical section: approved recommendation (1985) on IFCC methods the measurement of catalytic for concentration of enzymes. Part 3. IFCC method for alanine aminotransferase (I-alanine:2-oxoglutarate aminotransferase, ec 2.6.1.2). J Clin Chem Clin Biochem. 1986;24(7):481-95.
- Tietz NW, Rinker AD, 15. Shaw LM. International Federation Clinical of IFCC Chemistry. methods for the measurement of catalytic concentration of enzymes. Part 5. IFCC method for alkaline phosphatase (orthophosphoric-monoester phosphohydrolase, alkaline optimum, ec 3.1.3.1). IFCC document stage 2, draft 1, 1983-03 with a view to an IFCC recommendation. Clin Chim Acta. 1983; 135(3):339f-67f.
- Tietz NW, Rinker AD, Shaw LM. IFCC methods for the measurement of catalytic concentration of enzymes part 5. IFCC method for alkaline phosphatase (orthophosphoric-monoester phosphohydrolase, alkaline optimum, ec 3.1.3.1). J

Clin Chem Clin Biochem. 1983;21(11): 731-48.

- Tietz NW, Burtis CA, Duncan P, Ervin K, Petitclerc CJ, Rinker AD, et al. A reference method for measurement of alkaline phosphatase activity in human serum. Clin Chem. 1983;29(5):751-61.
- Gendler S. Uric acid. Kaplan A et al. Clin chem the CV Mosby Co St Louis Toronto Princeton. 1984;1268-73.
- Mihara M, Uchiyama M. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. Anal Biochem. 1978;86(1):271-8.
- 20. Marklund SL. Superoxide dismutase isoenzymes in tissues and plasma from New Zealand black mice, nude mice and normal balb/c mice. Mutat Res. 1985; 148(1-2):129-34.
- Abdel-bakky MS, Hammad MA, Walker LA, Ashfaq MK. Tissue factor dependent liver injury causes release of retinoid receptors (RXR-alpha and RAR-alpha) as lipid droplets. Biochem Biophys Res Commun. 2011;410(1):146-51.
- 22. Banchroft JD, Stevens A, Turner DR. Theory and practice of histoloicl techniques. Churchil Livingstone, New York, London, San Francisco, Tokyo; 1996.
- Ahsan R, Islam KM, Musaddik A, Haque E. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in albino rats. Global J Pharmacol. 2009;3(3):116-22.
- Kalu FN, Ogugua VN, Ujowundu CO, Nwaoguikpe RN. Aqueous extract of *Combretum dolichopentalum* leaf-A potent inhibitor of carbon tetrachloride induced hepatotoxicity in rats. Journal of Applied Pharmaceutical Science. 2011;1(10):114.
- 25. Dalton SR, Lee SM, King RN, Nanji AA, Kharbanda KK, Casey CA, et al. Carbon tetrachloride-induced liver damage in asialoglycoprotein receptor-deficient mice. Biochemical Pharmacology. 2009;77(7): 1283-90.
- Manibusan MK, Odin M, Eastmond DA. Postulated carbon tetrachloride mode of action: A review. J Environ Sci Health C Environ Carcinog Ecotoxicol Rev. 2007; 25(3):185-209.
- 27. Weber LW, Boll M, Stampfl A. Hepatotoxicity and mechanism of action of haloalkanes: Carbon tetrachloride as a

toxicological model. Crit Rev Toxicol. 2003;33(2):105-36.

- Jennings PE, Blandford RL, Rosenthal FD.
   Acute sulphasalazine hepatotoxicity.
   Postgrad Med J. 1986;62(726):305-6.
- 29. Posen S, Doherty E. The measurement of serum alkaline phosphatase in clinical medicine. Adv Clin Chem. 1981;22: 163-245.
- Dalton SR, Lee SM, King RN, Nanji AA, Kharbanda KK, Casey CA, et al. Carbon tetrachloride-induced liver damage in asialoglycoprotein receptor-deficient mice. Biochem Pharmacol. 2009;77(7):1283-90.
- Noori S, Rehman N, Qureshi M, Mahboob T. Reduction of carbon tetrachlorideinduced rat liver injury by coffee and green tea. Pakistan Journal of Nutrition. 2009; 8(4):452-8.
- 32. Tamayo RP. Is cirrhosis of the liver experimentally produced by cc14 an adequate model of human cirrhosis? Hepatology. 1983;3(1):112-20.
- Oakley F, Meso M, Iredale JP, Green K, Marek CJ, Zhou X, et al. Inhibition of inhibitor of kappa B kinases stimulates hepatic stellate cell apoptosis and

accelerated recovery from rat liver fibrosis. Gastroenterology. 2005;128(1):108-20.

- Azad Khan AK, Piris J, Truelove SC. An experiment to determine the active therapeutic moiety of sulphasalazine. Lancet. 1977;2(8044):892-5.
- Dooley S, Ten DP. Tgf-beta in progression of liver disease. Cell Tissue Res. 2012;347(1):245-56.
- Devaux Y, Bousquenaud M, Rodius S, Marie PY, Maskali F, Zhang L, et al. Transforming growth factor beta receptor 1 is a new candidate prognostic biomarker after acute myocardial infarction. BMC Med Genomics. 2011;4:83.
- Massague J, Chen YG. Controlling TGFbeta signaling. Genes Dev. 2000;14(6): 627-44.
- Devaux Y, Bousquenaud M, Rodius S, Marie PY, Maskali F, Zhang L, et al. Transforming growth factor beta receptor 1 is a new candidate prognostic biomarker after acute myocardial infarction. Bmc Med Genomics. 2011;4:83.
- Friedman SL. Evolving challenges in hepatic fibrosis. Nat Rev Gastroenterol Hepatol. 2010;7(8):425-36.

© 2016 Elsakka et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://sciencedomain.org/review-history/14979